### **Oxidative Stress**

in Cancer,
AIDS,
and
Neurodegenerative
Diseases

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# Nutriceutical Modulation of Glutathione with a Humanized Native Milk Serum Protein Isolate, IMMUNOCAL TM: Application in AIDS and Cancer

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## NUTRITIONAL IMMUNOMODULATION AND ITS RELATION TO GLUTATHIONE SYNTHESIS

Fresh, raw milk includes the group of proteins that remain soluble in "milk serum." These proteins can be preserved in their native form if extracted carefully from their natural source.

In 1981 it was discovered that normal mice fed a milk serum protein concentrate (specially prepared under mild nondenaturing conditions) exhibited a marked increase in the humoral immune response to a T helper cell-dependent antigen (1). In the following years, numerous experiments confirmed the consistency of this phenomenon (2-10). Over a period of 12 years and based on these findings a humanized native milk serum protein isolate (HNMPI) named Immuocal<sup>TM</sup> was developed (Immunotec Research Corporation Ltd., Montreal, Quebec, Canada).

This property was found to be related, at least in part to a greater production of splenic glutathione (L-.-glutamylcysteinylglycine) (GSH) during the oxygen-requiring antigendriven clonal expansion of the lymphocyte pool in animals fed with this bioactive HNMPI (9). Adequate levels of GSH are necessary for lymphocyte proliferation in the development of the immune response (11,12). Moderate but sustained elevation of cellular GSH as also found in the liver and the heart of healthy, old mice fed with this HNMPI for a prolonged period. In addition, HNMPI markedly increased their life expectancy in comparison to control animals fed nutritionally equivalent diets (13).

Glutathione is of major significance in cellular antioxidant activity in what Meister called the "GSH antioxidant system" because it participates directly in the destruction of reactive oxygen compounds and also because it maintains in reduced form ascorbate (vitamin C) and a-tocopherol (vitamin E), which also exerts a. antioxidant effect (14).

#### FUNCTION OF HNMPI AS A CYSTEINE DELIVERY SYSTEM

What ingredient in IMMUNOCAL<sup>TM</sup> makes it an effective "cysteine delivery system"? Systemic availability of oral GSH is negligible in man (I 5) and there is no evidence for transport of GSH into cells (16). Thus, it has to be synthesized intracellularly. This occurs in two steps: (a) glutamylcysteine synthesis; (b) glutathione synthesis. Even though the inflow of cysteine, glutamate, and glycine might prove somewhat limiting under selected circumstances, numerous observations have shown that it is the transport of cysteine (or cystine, which usually is promptly reduced to cysteine on cell entry) which tends to be the ate-limiting event in GSH synthesis. whereas free cysteine does not represent an ideal delivery system (17) because it is toxic and is spontaneously oxidized. Cysteine present as the disulfide cystine released during digestion in the gastrointestinal tract is more stable than free amino acid. GSH synthesis is submitted to negative feedback inhibition by the end-product GSH. The disulfide bond is pepsin- and trypsin-resistant, but may be split by heat and mechanical stress (9). Cystine accounts for about 90% of the low-molecular-mass cysteine in the blood plasma, while reduced cysteine is present only at extremely low concentration (18).

In a comparative study, we found that commercial milk serum concentrates exhibiting far less bioactivity, including less GSH promoting activity, contain about half the amount of serum albumin (9) and 4 times less lactoferrin than HNMPI, expressed as percentage of total milk serum protein. I..UNOCALTM is produced in a proprietary lenient process which results in the preservation of the most thermolabile proteins in their native conformation.

In the serum albumin, there are 17 cystine residues per 66 kDa molecule and 6 GluCys dipeptides (19); in lactoferrin there are 17 per 77kDa molecule and 4 Glu-Cys dipeptides (20); and in the @-lactalbumin there are 4 cystines in a 14,000 kDa molecule

Table 1.

	Molecular Mass (kDa)	<ti Residues</ti 	D=COLSPAN= 6 Cysteine Cysteine residues per molecule	Cysteine (Cys)2 (disulfide)	Glu-(Cys)2
P-Lactoglobuli	18,400	162	5	2	0
a-Lactabumin	14,200	125	8	4	0
Serum albumin	66,000	582	35	17	6
Lactoferrin	77,000	708	40	17	4

Source: Refs. 19, 20.

(19). On the other hand, P Lactoglobulin. has only 2 cystines in a 18,400kDa molecule (19), and IGGI, the predominant immunoglobulin in cow whey, has only 4 disulfide bridges in a 166,000kDa molecule (Table 1). In addition, Meister and colleagues (16) have demonstrated that the y-glutamylcysteine (Glu-Cys) precursors of GSH can easily enter the cell and there be synthesized into GSH. It thus become noteworthy that the most labile milk proteins-, serum albumin and lactoferrin-are those which contain these putative GSH-promoting peptide components.

Finally, the bioavailibility of the presumed active component (cystine and Glu-Cys group) may be influenced by the coexistence of the other proteins throughout the digestive-absorptive process.

This newly discovered property of HNMPI was found to be independent of its nutritional value, as other proteins of similar nutritional efficiency do not exhibit this unique property (1-10). The concept that a specific biological activity can exist in addition to and independent of the systemic effect Of IMMUNOCAL $^{\rm TM}$  as a good protein source is further substantiated by recent in vitro assays (21).

The dietary provision of cystine is particularly relevant to the immune system. The coordinated response of macrophages and lymphocytes in the T cell-mediated immune response is regulated, in part, by macrophage cystine uptake and subsequent release of reduced cysteine into the local environment for uptake by lymphocytes. When the antigenpresenting macrophages come into close contact with antigen-specific T cells, they supply these cells with additional amounts of cysteine and thereby raise their intracellular GSH level (18).

The validity of this assumption is confirmed by the demonstration that the immunoenhancing and GSH-promoting (data not shown) effect of Immunocal<sup>TM</sup> is abolished by buthionine sulfoximine, which inhibits y-glutamylcysteine synthetase, the initial step in GSH synthesis (17).

## IN VITRO MODULATION OF INTRACELLULAR GLUTATHIONE BY IMMUNOCAL TM

We demonstrated that normal human lymphocytes cultured for 3 days with HNMPI 100@g/ml show an increase in intracellular GSH content from  $4.5\pm0.4$  to  $10.5\pm3.4$ nmol/10 6 cells, p <0.01 (Figure 1). This increase in GSH correlates with an increase in cellular proliferation measured by thymidine incorporation (data not shown). The

**Table 2** Presence of Cytopathic Effects in MT-4 Cells

		TCID50/well3				
IMMUNOCAL <sup>TM</sup> (ug/ml)	2000	200	20	2		
0	+++	++	+	-		
I	+++	++	+	-		
10	++	+	+	-		
100	-	-	-	-		
500	-	-	-	-		
1000	-	-	-	-		

<sup>3+</sup> Presence of cytopathic effects; - absence of cytopathic effects.

increase in GSH is dose-dependent and has not been found for casein or for any commercially available milk serum protein concentrate (Figure 2).

#### IN VITRO ANTI-HIV and ANTIAPOPTOTIC ACTIVITY OF HNMPI

Clinically, there is direct evidence that HIV infection is associated ,with a GSH deficiency in the peripheral blood mononuclear cells (PBMC) (18). The depletion of intracellular GSH suggests an association between oxidative stress and HIV infection. Oxidative stress may be one of the mechanisms that contribute to disease progression and the wasting syndrome

through mediators of inflammation such as TNF-. and IL-6. During this period of progression, glutathione is consumed owing to an increase in oxidative stress.

GSH depletion, a consequence of chronic oxidative stress, is part of the spectrum of HIV infection. GSH has, in addition, a crucial role in lymphocyte function and cell survival. IMMUNOCAL<sup>TM</sup> functioning as a cysteine delivery system can enhance GSH synthesis in vitro (Figure 1) and inhibits HIV replication on a cord mononuclear cell system infected by HTL V-IIIB (Figure 3). IMMUNOCAL<sup>TM</sup> also inhibits the formation of syncitium between infected and noninfected cells. The inhibition of syncitium formation occurred at the same concentration as inhibition of HIV replication(Table 2). This viral inhibition was not associated with any cytotoxicity. IMMUNOCAL <sup>TM</sup>, via its GSH-promoting activity, educes apoptosis in HIV-infected cells. Apoptosis was evaluated by flow cytometry on PBMC from HIV-infected individuals (Dr. R. Olivier, AIDS and Retrovirus., Department, Pasteur Institute). HIV-infected PBMC cultured at concentrations of IMMUNOCAL' of I 00 ug/ml or higher were less prone to die of apoptosis than untreated cells:  $15\% \pm 2.6\%$  vs.  $37\% \pm 2.4$ , p<0.001 (Figure 4).

#### HNMPI SUPPLEMENTATION IN AIDS AND WASTING SYNDROME

Based on these preclinical data, we conducted a Canadian clinical trial (Canadian HIV Trials Network) with IMMUNOCAL $^{\rm TM}$  in children with AIDS and wasting syndrome. The major objective was to evaluate the effect of oral supplementation with IMMUNOCAL $^{\rm TM}$  on nutritional parameters and intracellular blood lymphocyte GSH concentration in children with AIDS and wasting syndrome. This was an open single-arm pilot study of 6 months duration. Wasting syndrome and severe weight loss within the 6 months preceding entry into the study was an absolute criterion for entry.

IMMUNOCAL<sup>TM</sup> was administered twice a day as a powder diluted in water. In some patients, IMNUNOCAL<sup>TM</sup> was administered via nasogastric tube when necessary. The administered starting dose was based on 20% of the total daily protein requirement and was increased by 5% each month over 4 months to reach 35% of the total protein intake at the end of the study. The total duration of the study was 6 months.

Weight, height, triceps skinfold and mid-arm muscle circumferences, CD4/CD8 counts, and peripheral lymphocyte GSH concentrations (measured by spectrophotometric assay) were measured monthly. Energy intake was assessed by the use of two independent 2-day food records with , 2-3 week period between the food records. Each food record included a weekday and a weekend, and the average of these records was calculated to reflect the daily nutritional intake. Out of 14 patients enrolled, 10 were evaluable. The ages of the patient were from 8 months to 15 years. The 10 patients studied were enrolled in four different centers across Canada: Montreal Children's Hospital (Dr. S. Baruchel), The Hospital For Sick Children Toronto (Dr. S. King), Children's Hospital for Eastern Ontario (Dr. U. Allen), and Centre Hospitalier Laval Quebec (Dr. F. Boucher). Of the 4 remaining patients, 2 lacked compliance after 2 months while the other 2 died of AIDS progressive disease within the first 2 months f entry into the study. None of the deaths was related to the tested product. >

None of the patients experienced any major toxicity such as diarrhea or vomiting or manifestation of milk intolerance. One patient had to Stop IMMUNOCAL $^{\rm TM}$  transiently for minor digestive intolerance such as nausea and vomiting (, twice/day) at month 3 and was subsequently able to restart the treatment without any problem.

At the end of the study, all patients experienced a weight gain in the range of 3.2% to 22% from their starting weight. The mean weight gain for the group was  $8.4\% \pm 5.7\%$ . On analysis of the mean percentage of requirement nutrient intake (RNI) per month for all

Table 3 Change from Baseline (expressed as a Percentage) at Weeks 24 and 36 in Weight, Anthropometric and GSH Patients Treated with IMMUNOCAL™

	Weight change%		Mid-arm Muscle circumference change		skinfold	Triceps skinfold change %		PBMC GSH change (%)	
Patient	wk 24	wk 32	wk 24	wk 32	wk 24	wk 32	wk 24	wk 32	
1	22.1	29.8	9.5	14.3	50.0	25.0	12.2	-9.0	
2	14.0	17.3	18.7	25.3	20.0	20.0	84.0	56.0	
3	5.1	9.2	-3.0	-2.0	-17.0	-3.0	37.0	55.0	
4	3.8	3.4	4.2	NA	-42.0	NA	305.0	550.0	
5	7.1	4.5	13.1	11.4	-24.0	-16.0	-18.0	14.3	
6	3.7	5.6	-2.0	-2.0	16.0	16.0	7.1	174.0	
7	2.5	NA	5.0	NA	-13.0	NA	54.2	NA	
8	14.2	18.2	-3.1	2.0	41.0	43.0	17.3	62.4	
9	8.9	7.9	-4.0	-8.0	-30.0	-39.0	-6.6	50.9	
10	7.0	NA	1.0	NA	41.0	NA	-1.6	NA	

#### NA

the patients, no correlation was found between the weight gain and any significant increase in the mean percentage of RNI, suggesting educed catabolism rather than an anabolic effect of IMMUNOCAL<sup>TM</sup>. Six of ten patients have demonstrated an improvement in their anthropometric parameters such as triceps skinfold or mid-arm muscle circumference independently of an increase in energy intake (Table 3).

Two groups of patients were identified in terms of GSH modulation: responders and nonresponders. The responders were those who started the study with a low GSH level.

The nonresponders were those who stated with a normal GSH level. A positive correlation was found between increase in weight and increase in GSH (Figures 5,6,7). No changes were found in terms of blood lymphocyte CD4 cell count, but 2 patients exhibited an

increase in the percentage of their CD8 cells and 4 patients showed a trend toward an increase in the number of NK cells.

In conclusion, this pilot study demonstrates that IMMUNOCAL<sup>TM</sup> is very well tolerated in children with AIDS and wasting syndrome and is associated FIGUREF an amelioration of the nutritional status of the patient as reflected by weight and antrhopometric parameters. Moreover, the GSH-promoting activity Of I.MUNOCAI<sup>TM</sup> in vivo seem to be validated in 6 out of 10 patients. An international multicenter double-blind randomized study is currently under way in France and Canada in adults patients with AIDS and wasting syndrome.

## SELECTIVE GLUTATHIONE MODULATION OF BREAST CANCER CELLS AND IMPACT ON CANCER CELL GROWTH

The specific involvement of GSH in the carcinogenic process is supported by the major role played by this compound in the detoxification of carcinogens by conjugation (26). We demonstrated that feeding GSH-promoting HNMPI to ice chronically treated with dimethylhydrazine (DMH) significantly reduces the number and size of colon carcinomas induced by DMH (27,28). These colon tumors appear to be similar to those found in the human insofar as the type of lesions and the chemotherapeutic response characteristics are concerned (26). HNMPI feeding appears to exert an inhibitory effect not only on the initiation (27) of cancer, but also on the progression of tumors (28).

Recently, a direct inhibitory effect of HNMPI in human cancer cell replication was confirmed (21,29,30). In other human cancer cell studies, the inhibitory effect ,as found to be related to the serum albumin component of milk serum (3 1) and most recently to @-lactalbumin (32). Feeding lactoferrin to mice inhibited the growth of solid tumors and in addition reduced lung colonization by melanomas (33). Unlike other proteins, serum albumin ,as found to exhibit a strong antimutagenic effect in an in vitro assay using hamster cells (34). It is therefore noteworthy that in this HNMPI we have succeeded in concentrating serum albumin, @-lactalbumin, and lactoferrin, all containing a significant number of GSH precursors. A possible explanation for these newly discovered properties of dietary milk serum protein may be found in recent findings on the role of GSH in tumor biology (35).

The search for ways to inhibit cancer cells without injuring normal cells has been based over the years on a vain effort to identify the metabolic parameters in which cancer cells are at variance with normal cells. One such function could well be the all-important synthesis of cellular GSH.

Recent experimental evidence has revealed an intriguing response of tumor versus normal cells to GSH synthesis-promoting compounds. Cellular GSH levels have been found to be several times higher in human cancer cells than in adjacent normal cells (35). This finding is presumably related to their proliferative activity. In fact, cancer is the only condition in which elevation of such a tightly regulated system as GSH has been reported. However, when a cysteine- and GSH-promoting compound such as 2-l-oxothiazolidine-4-carboxylate (OTZ) as added to cultured human lung cancer cells exhibiting very high levels of GSH at

the outset, no intracellular increase was noted, whereas GSH increased substantially in normal cells (35). This differential response is even more pronounced in vivo. We demonstrated that in tumor-bearing rats, OTZ treatment was actually found to deplete GSH in the tumors (36).

More specifically, an in vitro assay showed that, at concentrations that induce GSH synthesis and proliferation in normal human cells (Figure 1), IMMUNOCAL $^{\text{TM}}$  caused GSH depletion and inhibition of proliferation of cells in a rat mammary carcinoma (Figure 8) and Jurkat T cells (Figure 9) (21).

The selectivity demonstrated in these experiments may be explained by the fact that GSH synthesis is negatively inhibited by its own synthesis and since, as mentioned, baseline intracellular GSH in tumor cells is much higher than in normal cells, 'It is easier to reach the level at which negative feedback inhibition occurs in this cellular system than in a nontumor cellular system.

#### **HNMPI IN CANCER CLINICAL TRIALS**

On the basis of these experiments, 5 patients ,with metastatic carcinoma of the breast, I of the pancreas, and I of the liver, were fed 30 g of IMMUNOCAL<sup>TM</sup> daily for 6 months. In 6 patients, the blood lymphocyte GSH levels were substantially above normal at the outset, probably reflecting high tumor GSH levels. At completion of the 6 months of daily supplementation, 2 patients exhibited signs of tumor regression, normalization of hemoglobin and peripheral lymphocytes counts, and a sustained drop of lymphocyte GSH levels toward normal. Two patients showed stabilization of the tumor and increases in hemoglobin levels. In 3 patients, the disease progressed with a trend toward higher lymphocytes GSH levels (37).

A major problem in the use of chemotherapeutic agents in cancer therapy is the protection offered by the defense mechanisms of cancer cells. An important element of protection is represented by GSH, which is an effective detoxification agent that is relatively abundant in tumor cells. Indeed, when GSH synthesis is inhibited by buthionine sulfoximine (BSO), the activity of several chemotherapeutic agents such as alkylating agents is increased and drug resistance can be reversed (36-38). However, the concomitant depletion of GSH in normal cells greatly limits the practical usefulness of this modality of treatment. We recently demonstrated that a selective GSH prodrug such as OTZ protects some normal tissue (36) but also potentiates the activity of some alkylating agents (38). The apparently selective depletion of tumor GSH levels by provision of a natural precursor of GSH as contained in IMUNNOCALTM seems to be associated with inhibition of proliferation of cancer cells in vitro. This natural precursor of GSH favorably influences the GSH synthesis in normal cells. These in vitro and preliminary clinical results indicate that this newly discovered property of HNMPI may be a promising adjunct to the nutritional management of cancer patients undergoing chemotherapy. We are currently developing a phase 11 study in breast carcinoma, attempting to confirm that this selective depletion of GSH may, in fact, render tumor cells more vulnerable to chemotherapy and eventually protect normal tissue against the deleterious effect of chemotherapy.

#### ANALOGY BETWEEN HNMPI IMMUNOCALTM AND HUMAN MILK

Human milk contain about 80% of whey protein and 20% of casein. The opposite is true for cow milk. An analysis of the mass ratio of casein to whey protein in milk from various mammals clearly indicates that human milk has the lowest ratio in any mammalian species (39). On the basis of our laboratory studies showing the immunoprotective and anticancer effects of cow whey protein concentrate, it is tempting to speculate that this predominance of whey proteins in human milk is advantageous and thus represent an evolutionary adaptation.

Scientific data based on the similarity between the bioactive components of this native milk protein isolate (HNMPI) of cow milk, IMMUNOCAL<sup>TM</sup>, and human whey protein appear to substantiate this theory, as will now be discussed in more detail.

It is well known that breast feeding is superior to the use of cow milk-based formulas of similar nutritional efficiency for the health of human babies. Breast feeding protect against otitis media, and pneumonia (40,41). Mothers milk also has a protective effect on the incidence of several types of childhood cancer including leukemia, lymphomas, bone tumors, and brain tumors (42). Children who are artificially fed or are breast fed for only a shot period of time are more at risk for developing several types of cancer before the age of 15 years as compared to long-term breast feeders (43). Thus, the concept of a biological activity in addition to but independent of the nutritional efficiency, formulated to describe the immunoenhancing and GSH-promoting activity of the HNMPI IMMUNOCAL<sup>TM</sup>, may indeed apply to the breast feeding of neonates and infants. Glutathione synthesis appears to be the crucial factor in the health benefit of HNMPI.

It may then be appropriate to 'Identify the features common to HNMPI and human whey proteins that are capable of influencing GSH synthesis in the host. Cysteine, a crucial limiting factor in the synthesis of GSH, is about as abundant in cow's whey protein as it is in whole human milk proteins and several times more abundant than in cow's whole milk (39), since most caseins contain either no cysteine or one or two cysteine residues(19). As mentioned earlier, our studies showed that the most thermolabile milk proteins, namely, serum albumin, a-lactalbumin, and lactoferrin, are crucial to expression of the bioactivity of HNMPI. As shown in Table 1, these proteins are rich in cystine and glutamylcystine residues, natural precursors of GSH. The presence of these dipeptides in the product IMMUNOCAL<sup>TM</sup> is a characteristic shared with human milk (Table 4).

Traditionally, it has been advocated that "humanized" cow milk should contain more alactalbumin because this protein is twice as abundant in human milk. On the basis of our experimental findings, we propose instead that the principal health factor in human milk,

**Table 4.** Protein Composition of Cow and Human Milk Composition (g/liter)

Componet	Cow milk	Human milk	
Componer	COW IIIIK	Troman mink	(0 or 2 cysteine/molecule no disulfide bond)
Casein (g/l)	26	3.2	
B-Lactoglobulin(g/l)	3.2	Neglible	
a-Lactabumin (g/l)	1.2	2.8	
Serum albumin (g/l)	0.4	0.6	
Lactoferrin (g/l)	0.14	2.0	
Total cystine (mol/L)	8.19 x 10 -4	13.87 x 10 -4	
Total Cystine (mg/g protein)	6.4	38.7	

*Source:* Ref.19; Jennes R. Inter-species comaparison of milk proteins. In fox, ed. Developments in dairy chemistry-1. New York: ASP;1982:8

not denaturated by heat pasteurization, is due to the predominance of the thermolabile proteins rich in cystine and containing the Glu-Cys dipeptide which are characteristic of the bioactive HNMPI, namely, serum albumin, a-lactalbumin, and lactoferrin. This HNMPI differs from other commercially available milk serum protein concentrates in having a relatively high content of serum albumin (about I 0%), lactoferrin (about 0.65 %),

#### **CONCLUSION**

This article has addressed the central role of GSH in providing protection against endogenous oxiradicals and foreign pollutants. As an antioxidant, GSH is essential for allowing the lymphocyte to express its full potential, without being hampered by oxiradical accumulation during the oxygen-requiring development of the immune response. In a similar fashion, GSH delays the muscular fatigue induced by oxiradicals during the aerobic phase of strenuous muscular contraction.

It is, however, the second function of GSH-that of detoxification of chemical pollutants, carcinogens and ultraviolet radiation-that may well be of greater concern to medical science today, because of the ever-increasing demand on GSH as the major detoxifying agent. Under normal circumstances, a nutritionally balanced diet should provide sufficient precursors of GSH to allow for intracellular synthesis of adequate amounts of GSH. But in our current polluted environment, trace amounts of precursors found in an otherwise adequate diet may not be sufficient to allow for full GSH replenishment. This results in highly undesirable competition for GSH precursors developing amongst different systems. Cysteine prodrugs have helped clarify the essential role of GSH in athletic performance, immune function, AIDS, etc., but their effect is short-lived and their long-term use is not without adverse effects.

Using modern technology, it has been possible to obtain and consistently preserve, in their native form, the specific cow's milk proteins which share with predominant human milk proteins the same extremely rare GSH-promoting components. This product-the patented WPC-differs from most commercial WPCs in that it contains the active ingredients-notably cystine and glutamylcystine-in undenatured form and an amount sufficient to exhibit its potency when given as a dietary supplement, without overloading the system with excessive nitrogen intake.

It is therefore possible to obtain, with the patented milk serum protein concentrate, long-term moderate but sustained intracellular elevation of GSH and GSH precursors so that, when the challenge occurs, an efficient cellular response can be achieved.

#### REFERENCES

- 1. Meister A. The antioxidant effects of glutathione and ascorbic acid. In: Oxidative Stress, Cell Activation and Viral Infection. C. Pasquier et al (Eds.). Birkauser Verlag, Basel, Switzerland, 101-11, 1994.
  - 2. Meister A, Anderson ME. Glutathione. Ann Rev Biochem 52: 711-60, 1983.
  - 3. Kaplowitz N, Aw TY, Ookhtens M. The regulation of hepatic glutathione. Ann Rev Pharmacol Toxicol 25: 715-44, 1985. 4. Witschi A, Reddy S, Stofer B, Lauterburg BH. The systemic availability of oral glutathione. Eur J Clin Pharmacol 43: 667-9, 1992.
  - 5. Meister A. New aspects of glutathione biochemistry and transport selective alteration of glutathione metabolism. Nutr Rev 42: 397-410, 1984.
  - 6. Bounous G, Gold P. The biological activity of undenatured dietary whey proteins: role of glutathione. Clin Invest Med 14: 296-309, 1991.
  - 7. Droege W, Eck HP, Mihm S, Galter D. Abnormal redox regulation in HIV infection and other immunodeficiency diseases. In: Oxidative Stress, Cell Activation and Viral Infection. C. Pasquier et al (Eds). Birkauser Verlag, Basel, Switzerland, 285-99, 1994.
  - 8. Noelle RJ, Lawrence DA. Determination of glutathione in lymphocytes and possible association of redox state and proliferative capacity of lymphocytes. Biochem J 198: 571-9, 1981.
  - 9. Fidelus RK, Tsan MF. Glutathione and lymphocyte activation: A function of aging and auto-immune disease. Immunology 61: 503-8, 1987.
  - 10. Staal FJT, Roederer M, Israelski DM, Bubp J et al. Intracellular glutathione levels in T cell subsets decreases in HIV-infected individuals. AIDS Res and Hum Retro- viruses 8: 305-11, 1992.
  - 11. Herzenberg L, De Rosa S, Dubs G, Roederer M et al. Glutathione deficiency is associated with impaired survival in HIV disease. Proc Natl Acad Sci USA 94: 1967-72, 1997.
  - 12. Bounous G, Stevenson MM, Kongshavn PAL. Influence of dietary lactalbumin hydrolysate on the immune system of mice and resistance to Salmonellosis. J Infect Dis 144: 281, 1981.
  - 13. Bounous G, Kongshavn PAL. Influence of dietary proteins on the immune system of mice. J Nutr 112: 1747-55, 1982.
  - 14. Bounous G, Letourneau L, Kongshavn PAL. Influence of dietary protein type on the immune system of mice. J Nutr 113: 1415-21, 1983.

- 15. Bounous G, Kongshavn PAL. Influence of protein type in nutritionally adequate diets on the development of immunity. In: Absorption and Utilization of Amino Acids. M. Friedman (Ed.). Boca Raton, Florida: CRC Press, vol. 2, 219-32, 1989.
- 16. Bounous G, Batist G, Gold P. Immunoenhancing property of dietary whey protein in mice: role of glutathione. Clin Invest Med 12: 154-61,1989.
- 17. Bounous G, Shenouda N, Kongshavn PAL, Osmond DG. Mechanism of altered B-cell response induced by changes in dietary protein type in mice. J Nutr 115: 1409-17, 1985.
- 18. Hirai R, Nakai S, Kikuishi H, Kawai K. Evaluation of the Immunological Enhancement Activities of Immunocal. Otsuka Pharmaceutical Co. Cellular Technology Institute, Dec. 13, 1990.
- 19. Eigel WN, Butler JE, Ernstrom CA, Farrell HM et al. Nomenclature of proteins of cow's milk. Fifth revision. J Dairy Sci 67: 1599-631, 1984.
- 20. Goodman RE, Schanbacher FL. Bovine lactoferrin in RNA: Sequence, analysis, and expression in the mammary gland. Biochem Biophys Res Commun 180: 75-84, 1991.
- 21. Duncan B, Ey J, Holberg CJ, Wright AL et al. Exclusive breast-feeding for at least 4 months protects against otitis media. Paediatrics 91: 867-72, 1993.
- 22. Frank AL, Taber LN, Glezen WP, Kasel GL et al. Breast-feeding and respiratory virus infection. Paediatrics 70: 239-45, 1982. 23. Mather G, Gupta N, Mathur S, Gupta U. et al. Breast-feeding and childhood cancer. Indian Paediatrics 30: 652-7, 1993.
- 24. Davis MK, Savitz DA, Graubard BI. Infant feeding and childhood cancer. Lancet 1: 365-8, 1988.
- 25. Richie JP. The role of glutathione in aging and cancer. Exp Gerontol 27: 615-26, 1992.
- 26. Newberne PM, Butler WH. Acute and chronic effects of aflatoxins B1 on the liver of domestic and laboratory animals: A review. Cancer Res 29: 236-50, 1969.
- 27. Meerman JHN, Beland FA, Ketterer B, Srai SKF et al. Identification of glutathione conjugates formed from N-hydroxy-2-acetylaminofluorene in the rat. Chem Biol Interact 39: 149-68, 1982.
- 28. Boyland E, Sims P. The metabolism of benz(a)anthracene and dibenz(a,h)anthracene and their 5,6-dihydro derivatives by rat liver homogenates. Biochem J 97: 7-16, 1965.
- 29. Waterfall JF, Sims P. Epoxy derivatives of aromatic polycyclic hydrocarbons. The properties and metabolism of epoxides related to benzo(a)pyrene and to 7-8 and 9-dihydrobenzo(a)pyrene. Biochem J 128: 265-77, 1972.
- 30. Yamazoe Y, Roth RW, Kadlubar FF. Reactivity of benzidine diimine with DNA to form N-(deoxyguanosin-9-yl)-benzidine. Carcinogenesis 7: 179-82, 1986.
- 31. Bounous G, Papenburg R, Kongshavn PAL, Gold P et al. Dietary whey protein inhibits the development of dimethylhydrazine-induced malignancy. Clin Invest Med 11: 213-7, 1988.
- 32. McIntosh GH, Regester GQ, Le Leu RK, Royle PJ. Dairy proteins protect against dimethylhydrazine-induced intestinal cancers in rats. J Nutr 125: 809-16, 1995.
- 33. Frei E, Bertram B, Wiessler M. Reduced glutathione inhibits the alkylation by N-nitrosodimethylamine of liver DNA in vivo and microsomal fraction in vitro. Chem Biol Interact 55: 123-37, 1985.
- 34. Roberts JJ, Warwick GP. Mode of action of alkylating agents in formation of S-ethyl cysteine from ethyl methanesulphonate. Nature 179: 1181, 1958.
- 35. Coles B, Srai SKS, Waynforth B, Ketterer B. The major role of glutathione in the excretion of N, N-dimethyl-4-aminoazobenzene in the rat. Chem Biol Interact 47: 307-23, 1983.
- 36. Sims P. The metabolism of 3-methylcholanthrene and some related compounds by rat liver homogenates. Biochem J 98: 215-28, 1966.
- 37. Sims P. The metabolism of 7- and 12-methylbenz(a)anthra-cenes and their derivatives. Biochem J 105: 591-8, 1967.
- 38. Djuric Z, Coles B, Fifer EK, Ketterer B et al. In vivo and in vitro formation of glutathione conjugates from the K-region epoxides of 1-nitropyrene. Carcinogenesis 8: 1781-6, 1987.
- 39. Ripple MO, Henry W, Rago R, Wilding G. Prooxidant-antioxidant shift induced by androgen treatment of human prostate carcinoma cells. J Nat Cancer Inst 89: 40-8, 1997.
- 40. Hazelton GA, Lang CA. Glutathione contents of tissues in the aging mouse. Biochem J 188: 25-30, 1980.
- 41. Lang CA, Richie JP, Chen TS. Differential glutathione and cysteine levels in the brain of the aging mouse. Fed Am Soc Exp Biol, 1988. [Abstract 8327]
- 42. Lang CA, Naryshkin S, Schneider DL, Mills BJ et al. Low blood glutathione levels in healthy aging adults. J Lab Clin Med 120: 720-5, 1992.
- 43. Jeandel C, Nicolas MB, Dubois F, Nabey-Belleville F et al. Lipid peroxidation and free radical scavengers in Alzheimer's disease. Gerontology 35: 275-82, 1989.
- 44. Calvin HI, Medvedovsky C, Worgul BV. Near-total glutathione depletion and age-specific cataracts induced by

buthionine sulfoximine in mice. Science 28: 553-5, 1986.

- 45. Riederer P, Sofic E, Rausch WD, Schmidt B. Transition metals, ferritin, glutathione and ascorbic acid in Parkinsonian brains. J Neurochem 52: 515- 20, 1989.
- 46. Ebadi M, Srinivasan SK, Baxi MD. Oxidative stress and antioxidant therapy in Parkinson's disease. Prog Neurobiol 48: 1-19, 1996.
- 47. Kuzuya M, Naito M, Funaki C, Hayahi T et al. Protective role of intracellular glutathione against oxidized low density lipoprotein in cultured endothelial cells. Biochem Biophys Res Commun 163: 1466-72, 1989.
- 48. Bounous G, Gervais F, Amer V, Batist G et al. The influence of dietary whey protein on tissue glutathione and the diseases of aging. Clin Invest Med 12: 343-9, 1989.
- 49. Blumberg JB, Meydani SN. Role of dietary antioxidants in aging. In: Nutrition and Aging. Hutchinson MG, Munro HN (Eds.). New York: Academic Press, 85-97, 1986.
- 50. Birt DF, Baker PY, Hruza DS. Nutritional evaluations of three dietary levels of lactalbumin throughout the lifespan of two generations of Syrian hamsters. J Nutr 112: 2151-60, 1982.
- 51. Birt DF, Schuldt GH, Salmasi S. Survival of hamsters fed graded levels of two protein sources. Lab Anim Sci 32: 363-6, 1982.
- 52. Bray TM, Taylor CO. Enhancement of tissue glutathione for antioxidant and immune functions in malnutrition. Biochem Pharmacol 2113-23, 1994.
- 53. Puri RN, Meister A. Transport of glutathione, as g-glutamylcylsteinylglycyl ester, into liver and kidney. Proc Natl Acad Sci USA 80: 5258-60, 1983.
- 54. Anderson ME, Powric F, Puri RN, Meister A. Glutathione monoethyl ester: Preparation, uptake by tissues, and conversion to glutathione. Arch Biochem Biophys 239: 538-48, 1985.
- 55. Birnbaum SM, Winitz M, Greenstein JP. Quantitative nutritional studies with water-soluble, chemically defined diets. III. Individual amino acids as sources of "non-essential" nitrogen. Arch Biochem Biophys 72: 428-36, 1957.
- 56. Bridgeman MME, Marsden M, MacNee W, Flenley DC et al. Cysteine and glutathione concentrations in plasma and bronchoalveolar lavage fluid after treatment with N-acetylcysteine. Thorax 46: 39-42, 1991.
- 57. Williamson JM, Boettcher B, Meister A. Intracellular cysteine delivery system that protects against toxicity by promoting glutathione synthesis. Proc Natl Acad Sci USA 79: 6246-9, 1982.
- 58. Mant TGK, Tempowski JH, Volans GN, Talbot JCC. Adverse reactions to acetylcysteine and effects of overdose. Br Med J 289: 217-19, 1984.
- 59. Koch SM, Leis AA, Stokic DS, Khawli FA et al. Side effects of intravenous N-acetylcysteine. Am J Respir Crit Care Med 149: A321, 1994.
- 60. Williamson JM, Meister A. Stimulation of hepatic glutathione formation by administration of L-2-oxothiazolidine-4-carboxylate, a 5-oxo-L, prolinase substrate. Proc Natl Acad Sci USA 78: 936-9, 1981.
- 61. Baruchel S, Viau G, Olivier R, Bounous G. Nutriceutical modulation of glutathione with a humanized native milk serum protein isolate: Immunocal applications in AIDS and cancer. In: Oxidative Stress and Redox Regulation: Cellular Signaling, AIDS, Cancer and Other Diseases. Symposium May 21- 24, 1996, Institut Pasteur. [In press] 62. Watanabe A, Higuchi K, Yasumura S, Shimizu Y et al. Nutritional modulation of glutathione level and cellular
- 62. Watanabe A, Higuchi K, Yasumura S, Shimizu Y et al. Nutritional modulation of glutathione level and cellula immunity in chronic hepatitis B and C. Hepatology 24: 1883, 1996.